

## RESEARCH ARTICLE

# Bioactivity evaluation of organic solvent extractions of *Ganoderma lucidum*: a Sri Lankan basidiomycete

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**Abstract:** In this study, bioactivities of organic solvent extracts derived from nine species of terrestrial fungi were evaluated. The methanol extract of *Ganoderma lucidum* was selected as a potent sample with antibacterial activity showing growth inhibitory effects against pathogenic bacterial strains. Solvent-solvent partition chromatography was used to separate four fractions from the *G. lucidum* crude sample. Among the fractions, chloroform fraction was proved to comprise seven major compounds in HPLC analysis. The positive peak 4, at 27.0 min retention time on the UV detector at 290 and 365 nm, which corresponded to the negative peak at 680 nm on-line HPLC-ABTS<sup>+</sup> chromatogram was identified as an antioxidant compound. Antioxidant activity against 2,2-diphenyl-1-picrylhydrazyl (DPPH) was tested using radical scavenging activity with the electron spin resonance spectrometer (ESR). The highest activity was shown by the chloroform fraction with a IC<sub>50</sub> value 0.684 ± 0.31 mg mL<sup>-1</sup>. The inhibitory effect of nitric oxide (NO) production (%) was reported as the highest with the IC<sub>50</sub> value 30.38 ± 0.03 µg mL<sup>-1</sup> from chloroform fraction against lipopolysaccharide (LPS) induced RAW 264.7 cells *in vitro*. The methanol and ethyl acetate fractions also showed similar but lesser effects with IC<sub>50</sub> values 33.09 ± 0.04 and 122.9 ± 0.82 µg mL<sup>-1</sup>, respectively. Cell viability (%) by the MTT assay confirmed that there was no cytotoxic effect when treated with *G. lucidum* extracts at all concentrations. These results demonstrate the potential of using chloroform extracts of *G. lucidum* in antibacterial, antioxidant and anti-inflammatory therapeutic applications.

**Keywords:** Antibacterial activity, anti-inflammatory effect, antioxidant effect, *Ganoderma lucidum*, on-line HPLC-ABTS<sup>+</sup> terrestrial fungi.

## INTRODUCTION

Fungi are the major decomposers in certain ecosystems and essential associates of many organisms. High-throughput sequencing has estimated that there are more than 5.1 million fungal species in different habitats including soil, water and organisms (Blackwell, 2011). They are rich sources of secondary metabolites. Large numbers of fungal species are still chemically unexplored and conceived as promising sources of primary compounds for drug discovery and developments. The phylum basidiomycetes is one of the largest group of fungi among the four groups of true fungi; zygomycetes, basidiomycetes, ascomycetes, and deuteromycetes (Guarro *et al.*, 1999). Polypores are a large group of terrestrial fungi of the phylum basidiomycetes and a major source of pharmacologically active substances (Zjawiony, 2004).

The biochemical evaluation of terrestrial fungi has resulted in the isolation of a large number of secondary metabolites. Most of them are species specific and biosynthesized from a relatively small number of key intermediates, namely acetate, mevalonate, shikimate and amino acids. These metabolites are categorized based on their biosynthesis into groups such as terpenoids, polyketides, shikimates and alkaloids, and have shown pharmacological and medicinal value for the cure or alleviation of disease symptoms of humans. Therefore,

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mining of new bioactive secondary metabolites from terrestrial fungi is considered as an interesting aspect by many researchers recently.

Sri Lanka is a biodiversity hotspot comprising a high portion of endemic species among the fauna and flora: approximately 25 % of the flowering plants and 16 % of the mammals in the island are endemic. However, endemism among the 1920 fungal species of Sri Lanka is unknown. The earliest record of Sri Lankan fungi goes back to 1783; two species are recorded under the names *Peziza ceylonsche* and *Peziza limbosa* in "The Fungi of Ceylon" by Petch and Bisby (1950). Another publication by Coomaraswamy, (1979) gives a detailed account of the research carried out on Sri Lankan fungi and lists 2180 species belonging to 106 genera. *Ganoderma lucidum* and *Ganoderma applanatum* have been identified as the most common ceylon polypori occurring all over the island.

Only a limited number of chemical and biological studies have been done on Sri Lankan fungi. In a study to investigate the antibiotic activities of Sri Lankan fungi, 21 basidiomycetes species have been screened against bacterial strains including *Staphylococcus aureus*, *Streptococcus* sp., *Pseudomonas aeruginosa*, *Escherichia coli* and *Klebsiella* sp. *Volvariella volvacea* has been identified as a promising fungal species and led to the isolation of ergosterol peroxide (Perera et al., 2001). In addition, two new lanostane triterpenoids along with three known lanostanoids with anticancer activities have been isolated from *Ganoderma applanatum* (polyporaceae) (De Silva et al., 2006).

This study attempted to identify the prominent bioactive metabolites from randomly selected Sri Lankan terrestrial fungal species followed by the screening of their antibacterial activities. Antioxidant and anti-inflammatory potentials of organic solvent extracts of the prominent fungal species were determined using advanced chromatographic methods to evaluate their efficacy to isolate novel natural products.

## METHODS AND MATERIALS

### Chemicals and reagents

2,2-diphenyl-1-picrylhydrazyl (DPPH) and 2,2'-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid) (ABTS) were purchased from Sigma Chemicals Co. (St. Louis, USA). Nutrient agar (NA) and potato dextrose agar (PDA) were purchased from Difco (Sparks, MD, USA). All the chemicals and solvents used were of analytical grade.

### Fungi sample collection and extraction

Fresh samples of nine fungal species were collected based on the different morphological features of the fruiting bodies, from the Western Province of Sri Lanka during July and October 2007 and identified. Voucher specimens were deposited in the herbarium of the Department of Plant Science, University of Colombo, Sri Lanka. Freshly collected fruiting bodies of the fungal species were washed well and dried at room temperature for 24 h. The fruiting bodies were cut into small pieces and stored at -20 °C. These fungal samples were lyophilized and ground into fine powder. A 5 g quantity of each sample was mixed with 100 % methanol and kept in a shaking incubator at 25 °C for 1 day and filtered under a vacuum using a Whatman no.1 (Whatman Ltd., Maidenstone, England) filter paper. The methanol was evaporated in a rotary evaporator (Fisher Scientific, Loughborough, UK) and each of the sample was prepared into a 20 mg mL<sup>-1</sup> concentration.

### Disk diffusion assay

The methanol extracts from the terrestrial fungal species were screened for antibacterial activity using agar diffusion technique against 4 bacterial strains (Rota et al., 2004). The selected bacterial strains, *Bacillus subtilis*, *Staphylococcus aureus*, *Escherichia coli* and *Klebsiella aerogenes* were grown in nutrient agar. Sterilized filter paper disks (Whatman no. 1, 6 mm diameter) containing 50 µL (1 mg) of each extract was applied to the surface of the agar plates that were previously seeded with bacterial strains by spreading 0.1 mL of overnight culture. Methanol was used as a negative control. The plates were incubated overnight at respective temperatures, and the diameter of the growth inhibition zone was measured to the nearest millimeter. The scale of the measurement was as follows: > 9 mm clear zone as the strong inhibitory activity; 9-4 mm clear zone as the moderate/mild inhibitory activity; and < 4 mm as the low inhibitory activity, excluding the diameter of the filter paper disks (6 mm).

### On-line-HPLC analysis of *Ganoderma lucidum* extracts

Solvent-solvent partition chromatography was used to separate four different solvent extracts from *G. lucidum*. High-performance liquid chromatography (HPLC) system used in this experiment consisted of binary Waters 515 pump, Waters 2489 UV/Vis and 2998 photodiode array (PDA) detector and Waters 2707 auto sampler with the interface ABTS<sup>+</sup> radical analyser (Waters, Milford, MA 01757, USA). 10 µL of the 5 mg mL<sup>-1</sup> concentration

sample was run at 0.2 mL min<sup>-1</sup> flow rate on Atlantis T3 3 $\mu$ M 3.0  $\times$  150 mm column (Waters, USA) using a gradient of acetonitrile (ACN) ~ water solvent system (0 ~ 40 min: 10:100 v/v, 40 ~ 50 min: 100 % v/v) and the absorbance detected at 290 and 365 nm. For the preparation of 2,2'-azinobis(3-ethylbenzothiazoline-6-sulfonic acid) (ABTS) radical solution, 2 mM ABTS radical stock solution containing 2.5 mM potassium persulfate in water was diluted 30-fold using HPLC grade water. This solution was incubated overnight in dark at room temperature for radical stabilization. The separated analytes were sent to 'T' piece and reacted post-column with the ABTS radical in a reaction coil at 40 °C. The reduction was detected as a negative peak by UV detector set at 680 nm.

### DPPH radical scavenging assay

2,2-diphenyl-1-picrylhydrazyl (DPPH) is a free radical donor, which can be detected via electron spin resonance (ESR) spectrometry (JES-FA machine, JOEL, Japan) by the technique described by Nanjo *et al.* (1996). 60  $\mu$ L of each sample was added into 60  $\mu$ L of DPPH (60  $\mu$ mol L<sup>-1</sup>) in ethanol. After 10s of vigorous mixing, the solutions were transferred into 100  $\mu$ L Teflon capillary tubes and fitted into the cavity of the ESR spectrometer. The spin adduct was determined on an ESR spectrometer exactly 2 min later. The measurement conditions were as follows: central field 3475 G, modulation frequency 100 kHz, modulation amplitude 2 G, microwave power 5 mW, gain 6.3  $\times$  10<sup>5</sup>, temperature 298 K.

### Cell culture

The murine macrophage cell line RAW 264.7 was purchased from the Korean Cell Line Bank (KCLB, Seoul, Korea) and cultured in Dulbecco's modified eagle medium (DMEM) supplemented with 100 U mL<sup>-1</sup> of penicillin, 100  $\mu$ g mL<sup>-1</sup> of streptomycin and 10 % fetal bovine serum (FBS). The cells were incubated and maintained in an atmosphere of 5 % CO<sub>2</sub> at 37 °C and sub cultured every 2 d. Exponential phase cells were used throughout the experiments.

### Determination of nitric oxide (NO) production

RAW 264.7 cells (1  $\times$  10<sup>5</sup> cell mL<sup>-1</sup>) were placed in a 24-well plate and after 24 h, the cells were pre-incubated with various concentrations of the sample at 37 °C for 1 h. Further incubation was done for another 24 h with LPS (1  $\mu$ g mL<sup>-1</sup>) at the same temperature. After the incubation, the quantity of nitrite accumulated in the culture medium was measured as an indicator of NO production (Lee *et al.*, 2007). Briefly, 100  $\mu$ L of the cell culture medium was

mixed with 100  $\mu$ L of Griess reagent (1 % sulfanilamide and 0.1 % naphthylethylenediamine dihydrochloride in 2.5 % phosphoric acid), incubated at room temperature for 10 min, and the optical density at 540 nm was measured using an enzyme-linked immunosorbent assay (ELISA) microplate reader (Sunrise, Tecan Co. Ltd., Australia). The fresh culture medium was used as a blank in every experiment.

### Cytotoxicity assessment using MTT assay

The cytotoxicity of *G. lucidum* extracts against RAW 264.7 cell was determined using a colorimetric MTT assay. Cells were seeded in a 24-well plate at a concentration of 1  $\times$  10<sup>5</sup> cells mL<sup>-1</sup>. 24 hours after seeding, the cells were treated with extracts and incubated for an additional 24 h at 37 °C. MTT stock solution (50  $\mu$ L; 2 mg mL<sup>-1</sup> in PBS) was added to each well to a total reaction volume of 250  $\mu$ L. After 3 h of incubation, the supernatants were aspirated. The formazan crystals in each well were dissolved in 200  $\mu$ L of dimethylsulfoxide (DMSO), and the absorbance was measured with an ELISA plate reader at 540 nm.

### Statistical analysis

All the data were expressed as mean  $\pm$  standard deviation of three determinations. Statistical comparison was performed via a one-way analysis of variance followed by Duncan's multiple range test (DMRT). P values of less than 0.05 (p < 0.05) were considered as significant.

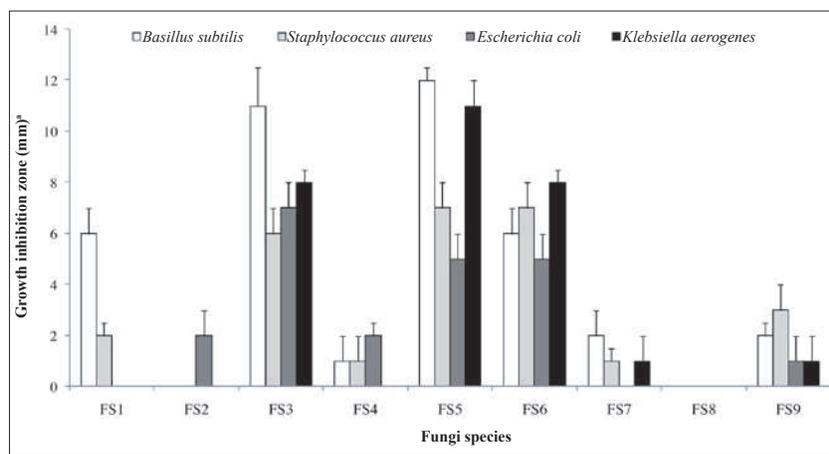
## RESULTS

### Antibacterial activity

The methanol extracts of nine randomly selected fungi were used to determine the growth inhibitory activity using the disc diffusion assay against four pathogenic bacterial strains including 2 Gram negative (*Escherichia coli* and *Klebsiella aerogenes*) and 2 Gram positive (*Bacillus subtilis* and *Staphylococcus aureus*) bacterial strains, respectively. The diameter of the growth inhibition zone (excluding the diameter of the filter paper disc - 6 mm) of pathogenic bacteria from the methanolic extracts of fungi are presented in Figure 1. Eight of the nine crude fungal extracts exhibited growth inhibition zones in at least one of the bacterial strains tested. Crude extracts of (FS3); *Daldinia cocentrica*, (FS5); *Ganoderma lucidum*, (FS6); *Pycnopoius cinnbarinus* and (FS9) *Xylaria polymorpha* showed a significantly high growth inhibition against all four pathogenic bacterial strains. Only the crude extract of *Trametes suaveolens* (FS8) showed no growth inhibition against all the pathogenic

bacterial strains. Among the growth inhibitory activity of crude extracts, *G. lucidum* showed the highest and promising antibacterial activity against all the Gram

positive and negative bacteria strains and was selected for further bioassays using different polar fractions after extraction from different organic solvents.



**Figure 1:** Diameter of the growth inhibition zone (excluded disk diameter of 6 mm) against the four pathogenic bacterial strains (*Bacillus subtilis*, *Staphylococcus aureus*, *Escherichia coli* and *Klebsiella aerogenes*) from the methanol extracts of the fungi species. The concentration of the fungi extract was used as 1 mg/disc.

<sup>a</sup> All data are means of three determinations

FS1: *Auricularia auricula judae*; FS2: *Agaric impudicus*; FS3: *Daldinia cocentrica*;

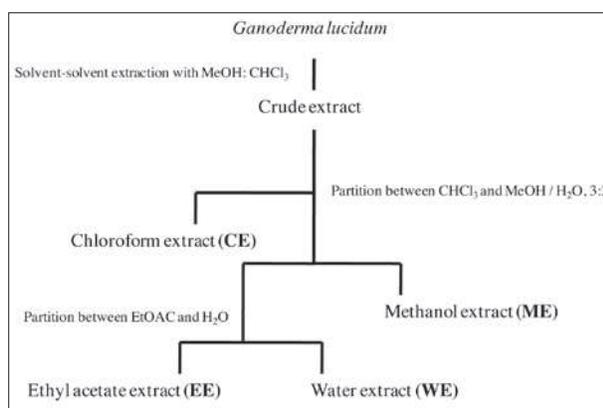
FS4: *Ganoderma* sp.; FS5: *Ganoderma lucidum*; FS6: *Pycnoporus cinnabarinus*;

FS7: *Schizophyllum commune*; FS8: *Trametes suaveolens*; FS9: *Xylaria polymorpha*

### *Ganoderma lucidum* extracts and on-line-HPLC profiles

The extracts of fruiting body of *G. lucidum* using (1:1) methanol:chloroform solvent system was obtained and

partitioned into different fractions according to the polarity of organic solvents used. The solvent-solvent partition chromatography approach (Figure 2) was used to fractionate the *G. lucidum* crude sample into four different fractions with varying polarity.



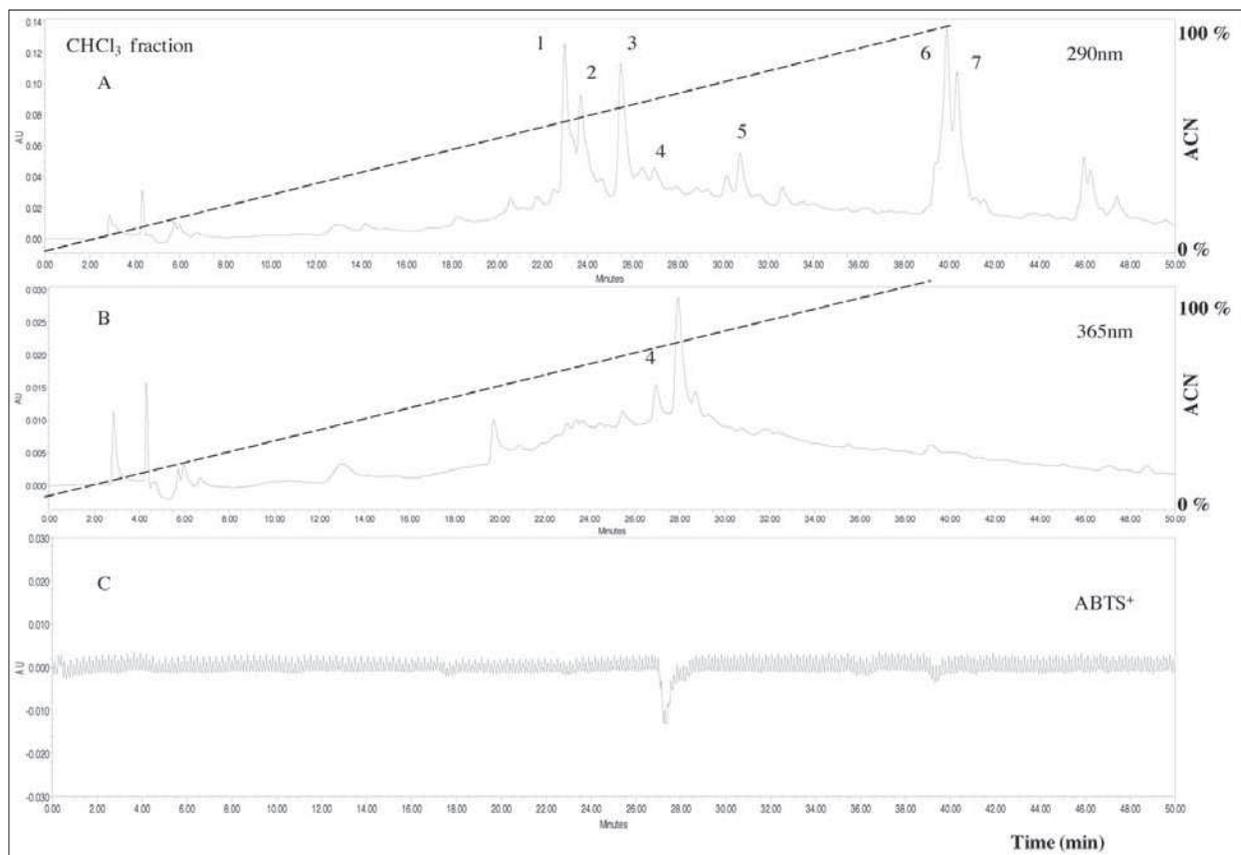
**Figure 2:** Extraction approaches of *Ganoderma lucidum* fungi sample using different organic solvents

Thus, methanol, chloroform, ethyl acetate and water fractions were used to analyze the on-line-HPLC profiles. Many compounds were found in the chloroform fraction only. Figure 3 (A) and (B) depict the chromatographic profile of the chloroform fraction of *G. lucidum* in the HPLC condition of a gradient of acetonitrile (ACN): water solvent system at flow rate 0.2 mL min<sup>-1</sup>(0 ~ 40 min: 10:100 v/v, 40 ~ 50 min: 100 % v/v) and absorption monitored at 290 and 365 nm, respectively. Seven major peaks were investigated in this study for chemical identification and only one peak (4) was determined to be of antioxidant value with the ABTS<sup>+</sup> radical scavenging effects. Figure 3 (C) depicts the ABTS<sup>+</sup> radical recorded at 680 nm as negative peak system. The positive peak (4) on the UV detector at 290 and 365 nm in on-line HPLC corresponded to the negative peak at 680 nm ABTS<sup>+</sup> radical chromatogram. The negative peak area

is dependent on the amount of antioxidant activity of the compound. The chloroform fraction gave positive evidence on the ABTS<sup>+</sup> radical scavenging activity and was considered as a fraction with potential for further purifications.

### Antioxidant activity

The scavenging activity of three different organic solvent extracts including methanol (ME), chloroform (CE) and ethyl acetate (EE) fractions of *G. lucidum* were measured



**Figure 3:** On-line HPLC chromatogram of the chloroform fraction from *Ganoderma lucidum*. (A) HPLC chromatogram of chloroform extract of *G. lucidum* at 290 nm and identified main compounds (1 – 7 peaks). (B) HPLC chromatogram of chloroform extracts of *G. lucidum* at 365 nm and identified antioxidative compound (peak 4). (C) On-line HPLC-ABTS<sup>+</sup> chromatogram of chloroform extract of *G. lucidum* with ABTS radical reagent and the negative peak with respect to peak 4, indicating the antioxidant activity determined at 680 nm.

against DPPH free radicals using the electron spin resonance spectrophotometer (Figure 4). The chloroform fraction showed the highest DPPH radical scavenging activity with an IC<sub>50</sub> value of 0.684 ± 0.31 mg mL<sup>-1</sup>. The methanol and ethyl acetate fractions showed IC<sub>50</sub> values as 1.109 ± 0.54 and 2.09 ± 0.42 mg mL<sup>-1</sup>, respectively. All fractions increased the DPPH radical scavenging activity (%) with the increase of concentration, in a dose dependent manner.

### Anti-inflammatory activity in terms of NO production inhibitory effect

Nitric oxide (NO) production by the inflammatory mediators by iNOS, with the different *G. lucidum* fractions were determined in LPS-activated RAW 264.7 cell *in vitro*. LPS acts as endotoxins for mammals and stimulates the RAW cells in terms of enhancing the NO concentration in the medium. However, with the

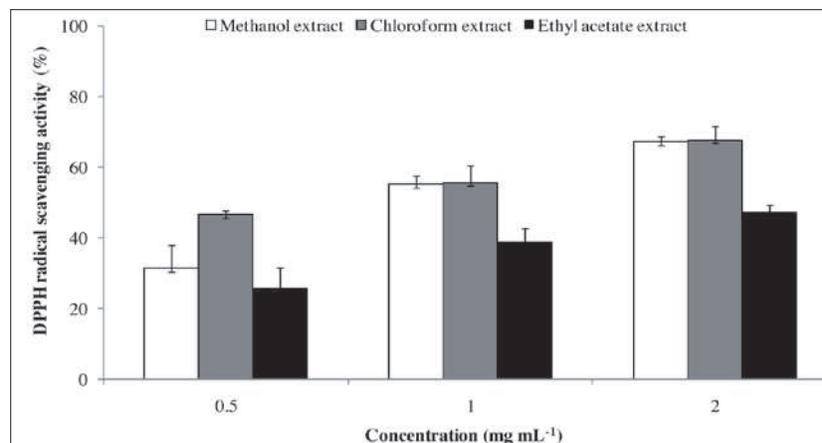
pretreatment of extracts, decrease of NO production level (%) was measured at all the concentrations (Figure 5). In this study, methanol and chloroform fractions showed the highest and similar results for inhibitory effect of NO production (%) on LPS-induced RAW macrophages dose dependently. The calculated  $IC_{50}$  values on ME and CE fractions were  $33.09 \pm 0.04$  and  $30.38 \pm 0.03 \mu\text{g mL}^{-1}$ , respectively (Table 1). The EE fraction showed a lower activity with  $IC_{50}$  values  $122.9 \pm 0.82 \mu\text{g mL}^{-1}$ . Therefore, the determined NO production inhibitory activity (%) was about 4-fold lesser in EE fraction than in ME and CE fractions. In addition, the cytotoxic effects on RAW 264.7 cells with the treated samples were performed by MTT assay. In these tests, *G. lucidum* fractions did not show a significant cytotoxic effect at all treated concentrations. Therefore, *G. lucidum* extracts can be considered as potential agents for suppressing NO production without any cytotoxic effect for anti-inflammatory activity.

## DISCUSSION

Sri Lankan basidiomycetes are rich sources of secondary metabolites and preliminary screening bioassays might

help to identify possible bioactivities. Nine species of fungi randomly selected from Sri Lankan habitats were tested for antibacterial activity by disc diffusion assay against 2 Gram negative (*Escherichia coli* and *Klebsiella aerogenes*) and 2 Gram positive (*Bacillus subtilis* and *Staphylococcus aureus*) bacterial strains, respectively. The highest growth inhibition activity was reported in the *G. lucidum* sample against all the bacterial strains. *G. lucidum* is a well-known terrestrial fungus and occurs in most of the countries. Some countries including China, Korea and Japan are using *G. lucidum* extracts or fresh materials directly or indirectly for treatment in traditional medicine (Yue *et al.*, 2010). Recent studies have discussed the pharmacological and medicinal value of *G. lucidum*. This is the first study on Sri Lankan *G. lucidum* that has evaluated and reported the bioactivities against antibacterial, antioxidant and anti-inflammatory effects.

In this study, extracted organic solvent fractions were tested using HPLC and the chloroform fraction that had the highest amount of possible secondary metabolites was identified. The on-line HPLC-ABTS<sup>+</sup> chromatogram

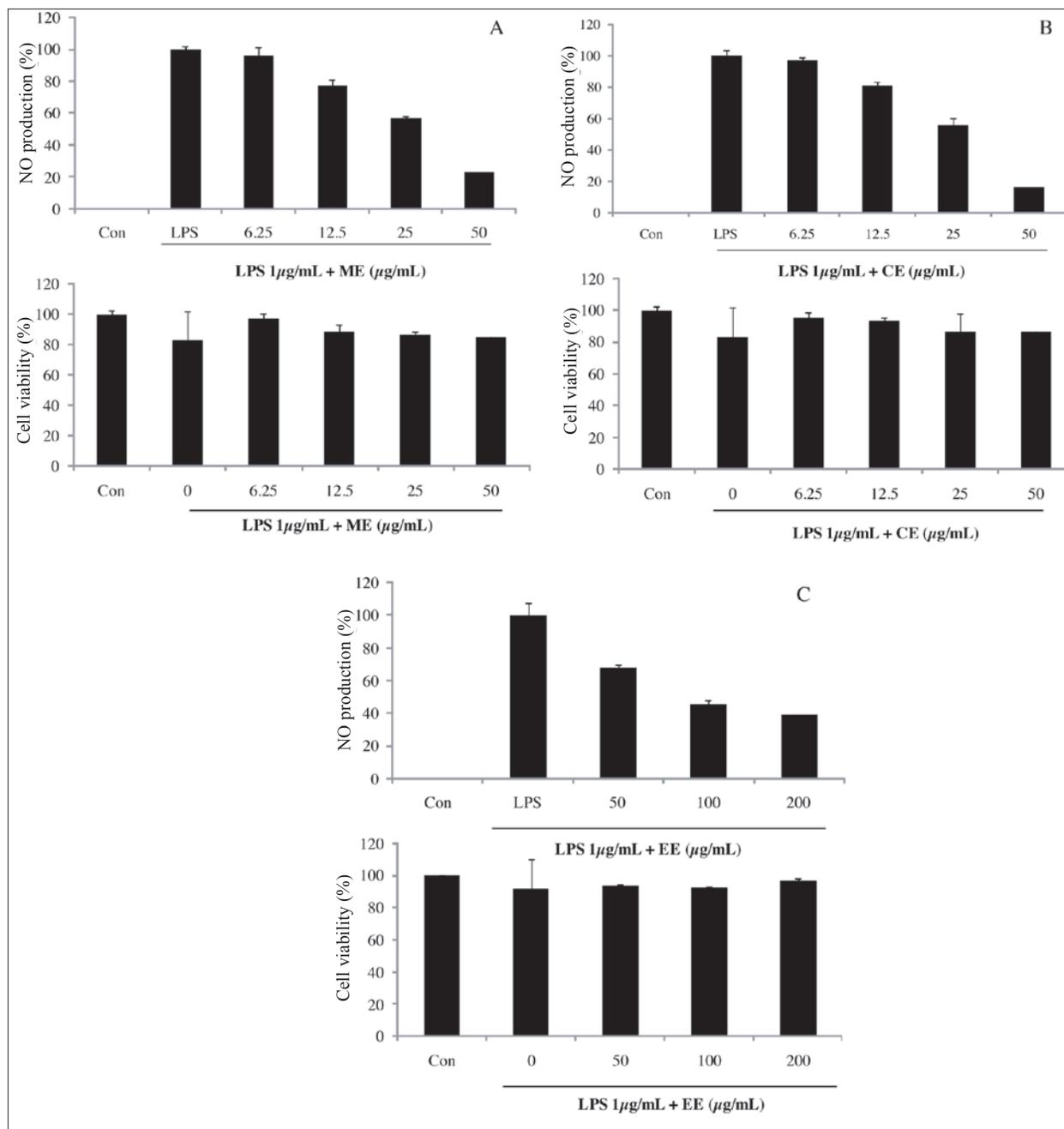


<i>Ganoderma lucidum</i> extracts			
DPPH Radical scavenging activity (%)	ME	CE	EE
$IC_{50}$ (mg mL <sup>-1</sup> ) <sup>a</sup>	$1.109 \pm 0.54$	$0.684 \pm 0.31$	$2.09 \pm 0.42$

**Figure 4:** DPPH radical scavenging activity (%) of the organic solvent extracts of *Ganoderma lucidum* determined using electron spin resonance spectrometer (ESR).

<sup>a</sup> The concentration of the extract required to scavenge 50 % of the antioxidant activity. Values of  $IC_{50}$  were determined by the triplicate individual experiments and values are mean  $\pm$  SD of three determinations.

ME: methanol extract, CE: chloroform extract, EE: ethyl acetate extract.



**Figure 5:** NO production inhibitory effect of *Ganoderma lucidum* organic solvent extracts; (A): methanol extract; ME, (B): chloroform extract; CE and (C): ethyl acetate extract; EE on lipopolysaccharide (LPS) induced NO production (%) and cell viability (%) in RAW 264.7 macrophages. After incubation of the extract with cells in response to LPS ( $1 \mu\text{g mL}^{-1}$ ) for 24 h, the NO levels in the medium was measured. Values are mean  $\pm$  SD of three determinations.

profile has shown 7 major compounds in the chloroform fraction. Interestingly, the negative peak at 680 nm of ABTS<sup>+</sup> profile at 27.0 min retention time (UV detector

at 290 and 365 nm) was identified as the antioxidant compound from the chloroform fraction of *G. lucidum* extract.

**Table 1:** Anti-inflammatory activity of *Ganoderma lucidum* extracts on LPS-induced RAW 264.7 macrophages

NO production inhibitory activity (%)	<i>Ganoderma lucidum</i> extracts		
	ME	CE	EE
IC <sub>50</sub> (µg mL <sup>-1</sup> ) <sup>a</sup>	33.09 ± 0.04	30.38 ± 0.03	122.9 ± 0.82

<sup>a</sup> The concentration of the extract required to scavenge 50 % of the NO production inhibitory activity (%). Values of IC<sub>50</sub> were determined by the triplicate individual experiments and values are mean ± SD of three determinations. ME: methanol extract, CE: chloroform extract, EE: ethyl acetate extract

Following the separation of chloroform extract at 5 mg mL<sup>-1</sup> concentration on the HPLC column, the elute was directed to PDA detector and mixed with a stabilized solution of ABTS<sup>+</sup> radicals. ABTS<sup>+</sup> radicals are quenching and the results indicate a negative peak on the HPLC trace as illustrated in the A<sub>680 nm</sub> profile (Figure 3). The area under the peak depends on the strength of antioxidant activity and is described as an advanced technology to describe natural products quantification through the on-line HPLC-ABTS<sup>+</sup> chromatogram (Stalmach *et al.*, 2006). In addition, DPPH radical scavenging activity (%) showed that the chloroform fraction has a higher activity (IC<sub>50</sub> value 0.684±0.31 mg mL<sup>-1</sup>) compared to the other organic solvent extracts of *G. lucidum*. In this study, an advanced methodology was used to detect the radical scavenging activity, using electron spin resonance spectrophotometer (ESR). Spin trapping is the most direct method for the detection of highly reactive free radicals, which can overcome the sensitivity problem inherent in the detection of endogenous radicals in biological systems. The prominent antioxidant value achieved from the chloroform fraction complied with the on-line HPLC-ABTS<sup>+</sup> chromatogram.

Inflammation is one of the physiological processes initiated due to the pathogenic invasion or cell and tissue injury (Wadleigh *et al.*, 2000). This can be influenced by the activation of various immune cells such as macrophages, neutrophils and lymphocytes. Nitric oxide (NO) plays an important role in many cellular functions in the biological systems as a signalling molecule (Wang *et al.*, 2006). Importantly, NO is generated in macrophages as a part of the human immune responses (Kassim *et al.*, 2010). Therefore, inflammatory mediators such as nitric oxide (NO) and prostaglandin (PGE<sub>2</sub>) are also activated due to inducible nitric oxide synthase (iNOS) and cyclooxygenase-2 (COX-2). In this study, the pathological condition was stimulated by lipo-polysaccharides (LPS) and the NO production is

increased by iNOS. The suppression of iNOS expression and inhibition of NO production by the *G. lucidum* samples were determined to be of therapeutic potential value of anti-inflammation. The profound inhibitory effect observed in the chloroform extracted fraction (IC<sub>50</sub> value 33.09 ± 0.04 µg mL<sup>-1</sup>) and cytotoxic effect against RAW 264.7 macrophages was not seen at all the concentrations. According to the results, the chloroform extract of *G. lucidum* has profound bioactivities including antibacterial, antioxidant and anti-inflammatory effects. Furthermore, the literature survey has supported the fact that triterpenoid compounds are contained in the chloroform fraction of *G. lucidum* resulting in bioactivities. Yang *et al.* (2007) have reported that the chloroform fraction of *G. lucidum* separated a total of 32 triterpenoids, recently. The chloroform fraction of the Sri Lankan *G. lucidum* has shown profound antibacterial, antioxidant and anti-inflammatory activities, which might have potential to isolate unknown and interesting triterpenoid structures.

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