

RESEARCH NOTE

Chemistry of *Heterodermia microphylla*, a lichen new to Sri Lanka

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Lichens are biologically distinct symbiotic entities composed of an algal or cyanobacterial photobiont and a fungal mycobiont¹. Lichens synthesize a great variety of metabolites with different structures and potential biological activities^{2,3}. In our continuing research on the chemistry of tropical lichens, we have investigated a foliose type lichen belonging to the family Physciaceae, *Heterodermia microphylla* (Kurok.) Skorepa (Figure 1), a lichen which has not been reported from Sri Lanka prior to this study. The thallus is appressed and marginally ascendant; its upper surface, and sometimes both surfaces, may contain a cortex⁴. The lower surface is corticated with rhizinae which are found only along the margin⁵. The photobiont of *Heterodermia* is a green alga, which is a chlorococcoid. The thallus of *H. microphylla* has lobes slightly disjunct or adjunct, more or less plane, not ascending, with short lateral lobes⁶. Underside of the thallus is non-corticate and pale. *H. microphylla* is present in Asia and New Zealand. This lichen can be found at altitudes of 1500-2300 m growing on tree trunks

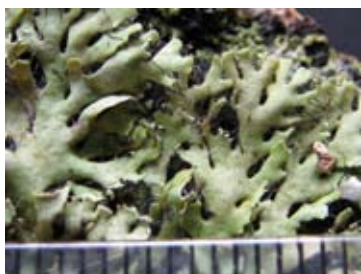


Figure 1: A part of the thallus (upper surface) of *H. microphylla*

in sheltered but open woodland and occasionally on rock. The genus *Heterodermia* is useful in determining the ecological continuity of forest ecosystems⁷.

H. microphylla growing on a rock beside a small stream was collected from Labukelle, Central Province. Dried, cleaned specimens of *H. microphylla* (26.05 g) were sequentially extracted with 2.5 L of hexane and CH₂Cl₂ (2.5 L).

The hexane extract (242 mg) when subjected to flash chromatography (eluent: 50% hexane/CH₂Cl₂) yielded the following: Methyl-5-chloro-3-formyl-2,4-dihydroxy-6-methylbenzoate (**1**): 6mg⁸; Methyl-β-ornicincarboxylate (**2**): 8 mg; m.p. 144-143 °C, lit.⁹ 140-141 °C; Zeorin (**3**): 42 mg; m.p. 236-242 °C lit.¹⁰ 240-243) °C.

The CH₂Cl₂ extract (460 mg) when subjected to Medium Pressure Liquid Chromatography (MPLC) (hexane to CH₂Cl₂) followed by flash chromatography (eluent: 1% MeOH/CH₂Cl₂) yielded: Atranorin (**4**): 43 mg; m.p. 194-196 °C, (lit m.p.¹¹ 196 °C); and Chloroatranorin (**5**): 6 mg; m.p. 207-210 °C (lit. m.p.¹² 208-208.5 °C).

Upon close examination, the Nuclear Magnetic Resonance (NMR) data (¹H and ¹³C) of compound (**1**) were identical to methyl chlorohematommate, isolated from the lichen *Lethariella canariensis*, its only other reported source in nature⁸. The hexane extract also yielded methyl-β-ornicincarboxylate (**2**) and the triterpenoid zeorin (**3**). In addition to chloroatranorin (**5**), the CH₂Cl₂ extract yielded atranorin (**4**). The latter

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exhibited moderate larvicidal activity against the 2nd instar larvae of *Aedes aegypti* (90 and 100 % mortality at 100 ppm after 24 and 48 h, respectively).¹³

The Thin Layer Chromatography (TLC) bioassay¹⁴ indicated the antifungal activity of compound (4) against *Colletotrichum gloeosporioides*. A quantitative spore germination assay¹⁵ corroborated the above result (Table 1) indicating that atranorin (4) was comparable in activity to the standard benlate (methyl-1-(butylcarbonyl)-2-benzamidazolecarbamate) against *C. musae*.

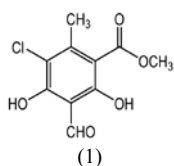
From a chemotaxonomic point of view, this is the first report of the occurrence of compound 1 in the genus *Heterodermia*. and the family Physiaceae.

Table 1: Spore germination results for atranorin

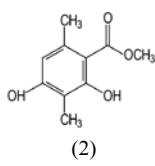
Fungus	Compound	% Germination
<i>C. gloeosporioides</i>	4 ^a	58.2 ± 3.3
	control ^b	74.2 ± 1.4
	benlate ^a	33.3 ± 1.1
<i>C. musae</i>	4	26.2 ± 2.3
	control	66.8 ± 1.1
	benlate	32.4 ± 3.3

^a Concentration of both compound 4 and benlate was 10 ppm

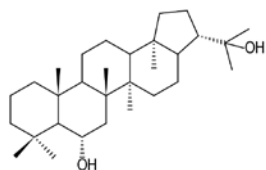
^b Negative control with no compound



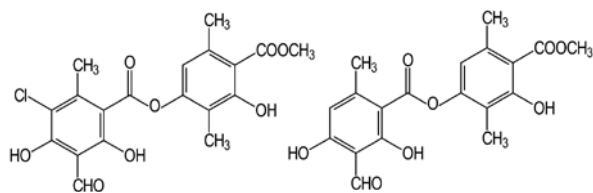
(1)



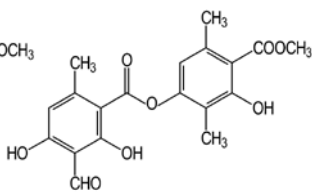
(2)



(3)



(4)



(5)

Acknowledgement

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References

- Nash T.H. III. (1996). *Lichen Biology*. Cambridge University Press, U.K.
- Huneck S. (1999). The significance of lichens and their metabolites. *Naturwissenschaften* **86**: 559-570.
- Huneck S. & Yoshimura I. (1996). *Identification of Lichen Substances*. pp. 113-123. Springer, Berlin.
- Kurokawa S. (1962). A monograph of the genus *Anapychia*. *Nova Hedwigia* **6**: 1-115.
- Kurokawa S. (1973). Supplementary notes on the genus *Anapychia*. *Journal of the Hattori Botany Lab* **37**: 563-607.
- Swinscow T.D.V. & Krog H. (1988). *Macrolichens of East Africa*. pp. 97-98. British Museum (Natural History), London.
- Coppins A.M. & Coppins B.J. (2002). *Indices of Ecological Continuity for Woodland Epiphytic Lichen Habitats in the British Isles*. pp. 1-36. British Lichen Society UK.
- Marante F.J.T., Castellano A.G., Rosas F.E., Aguiar J.Q. & Barrera J.B. (2003). Identification and quantitation of allelochemicals from the lichen *Lethariella canariensis*: phytotoxicity and antioxidative activity. *Journal of Chemical Ecology* **29**(9): 2049-2071.
- Hylands P.J. & Ingo-lfsdo-ttir K. (1985). The isolation of methyl β-orsellinate from *Stereocaulon alpinum* and comments on the isolation of 4,6-dihydroxy-2-methoxy-3-methylacetophenone from *Stereocaulon* species. *Phytochemistry* **24**: 127-129.
- Yosioka I., Nakanishi T., Yamaki M. & Kitagawa I. (1972). The final conclusion on the stereostructure of zeorin and its correlation with leucotylin. The structure of isoleucotylin. *Chemical and Pharmacological Bulletin* **20**: 147-156.
- Brasy P.C., Bachet B., Bodo B. & Molho D. (1982). Structure de l'atranorine. *Acta Crystallography Section B* **29**: 889-890.
- Culberson C.F. (1969). *Chemical and Botanical Guide to Lichen Products*. p.p. 141-145. University of North Carolina Press, Chapel Hill, USA.
- Bandara K.A.N.P., Kumar V., Jacobsson U. & Molleyres P. (2000). An insecticidal piperidine alkaloid from *Microcos paniculata* stem bark. *Phytochemistry* **54**: 29-32.
- Klarman W.L. & Stanford J.L. (1968). Isolation and purification of an antifungal principle from infected soybeans. *Life Sciences* **7**: 1095-1103.
- Anonymous (1943). *Slide germination for evaluating fungicides*. *American Phytopathological Society, Committee on standardization of fungal tests* **33**: 627.