

SHORT COMMUNICATION

Cytotoxicity and antioxidant activity studies of green leafy vegetables consumed in Sri Lanka

B. M. G. K. Balasuriya* and H. R. W. Dharmaratne

Natural Products Programme, Institute of Fundamental Studies, Hantana Road, Kandy.

Revised: 21 February 2007 ; Accepted: 04 September 2007

Abstract: This investigation was conducted to examine the cytotoxicity and the antioxidant activity of some green leafy vegetables consumed in Sri Lanka. In this study, cytotoxicity was tested using brine shrimp (*Artemia salina*) lethality bioassay. As anticipated majority of the tested leafy vegetables were found to have insignificant cytotoxicity. However, some of the greens *Aerva lanata* and *Baccopa monnieri* showed significantly higher level of ($p < 0.001$) cytotoxicity when compared with the positive control. *Alternanthera sessilis* which is the most popular leafy vegetable among Sri Lankans and *Passiflora edulis* showed similar toxicity levels as the positive control. Consumption of these four leafy vegetables could pose a potential health risk. Therefore, further toxicological studies should be carried out to evaluate their potential health risks. Antioxidant activity of above greens were tested using 2,2-diphenyl-1-picrylhydrazyl (DPPH) assay, and all the tested leafy vegetables showed free radical scavenging properties indicating the presence of primary antioxidants in the plants.

Key words: Antioxidant activity, *Artemia salina*, cytotoxicity, DPPH, green leafy vegetables

INTRODUCTION

Green leafy vegetables (greens) play a significant role in the Sri Lankan diet probably due to the influence of traditional herbal medicine, their low cost and easy availability. Further, greens are a major source of vitamins, minerals and fiber. Due to scientific or other reasons, the use of certain herbs in the diet has been discontinued while others have been added. Information as well as misinformation plays a major role in this process. The removal of nutritional herbs from the diet based on inadequate or erroneous information denies access to an easily available cheap source of nutrients while use of herbs about which little is known may endanger human health. There are reports worldwide

about misidentification of greens and culinary herbs, leading to the consumption of toxic herbs which can cause irreparable damage to organs and body function. Hence, there is a vital need to educate the general public on their choice of green leafy vegetables for consumption, which necessitates a systematic scientific evaluation of their nutritional properties as well as the toxic effects.

Antioxidants exert a protective role in cancer and tumor prevention processes in the human body by quenching oxidation type free radicals in blood. Therefore, it is essential to consume foods rich in antioxidants. The present investigation was focused mainly on cytotoxicity and antioxidant activity studies of popular greens and culinary herbs used in Sri Lanka.

METHODS AND MATERIALS

Thirty types of pesticide free green leafy vegetables were collected mainly from the field. Cleaned plant material (100 g) was macerated in distilled water (100 mL), filtered through a muslin cloth and filtrate was freeze-dried to get the water extract.

Brine shrimp (*Artemia salina*) lethality bioassay^{1,2} was used for the preliminary screening for cytotoxicity of water extracts of the leafy vegetables, as this assay has shown a good correlation with cytotoxicity assays with human cell lines and mouse bioassays¹. Brine shrimp, *Artemia salina* eggs (imported from Pets Food, Sam Yu Foods Co.Ltd, Taiwan) were purchased from the local pet shop. Eggs were hatched in artificial sea water² at room temperature (27 °C) and aerated for 48 h. Second instar nauplii were used in the test since they have been reported to show the highest sensitivity³ to test substance.

* Corresponding author

Tests were performed in the wells of 96-well plates in triplicates of 1000, 500, 100 and 10 ppm concentrations of the 30 freeze-dried water extracts of selected greens. Dimethyl sulfoxide (DMSO): seawater, 1:19 was used as the solvent. When the extract is water soluble, seawater was used as the solvent. DMSO: seawater ratio was determined by a separate experiment performed to detect the ineffective DMSO dilution. Ten 2nd instar nauplii were added to each well of 96-well plates and the number of survivors was counted after 24 h of incubation at room temperature (27 °C). Quinoline in the solvent was used as the positive control, while negative control was performed by adding only the solvent. The test was repeated three times. LC₅₀ values were calculated using

EPA probit analyzer version 1.5. The data were shown as mean ± SE. One-way ANOVA with Fisher's comparison was adopted to evaluate the statistical significance. Probability level was fixed to p<0.05 (Table 1).

2,2-diphenyl-1-picrylhydrazyl (DPPH) free radical scavenging assay was used for the preliminary screening of anti-oxidant activity⁴. Freeze-dried water extracts of green leafy vegetables and tocopherol (positive control) were spotted on thin layer chromatography (TLC) plates, and exposed to a free radical reagent 0.4M ethanolic solution of DPPH to visualize anti-oxidant activity. Antioxidants neutralize the purple colour DPPH free radicals in to bright yellow colour.

Table 1: Cytotoxicity of different vegetable greens on brine shrimps

Botanical name	Vernicular name	LC ₅₀ value (ppm)
<i>Alternanthera sessilis</i>	Mukunuwana	185.31 ± 0.01 ^A
<i>Centella asiatica</i>	Gotukola	793.33 ± 1.81
<i>Murraya koenigii</i>	Karapincha	810.45 ± 8.37
<i>Brassica sativa</i>	Kola goa	308.80 ± 13.1
<i>Ipomoea aquatica</i>	Kankun	385.02 ± 1.21
<i>Basella alba</i>	Nivithi kola	>1000
<i>Passiflora edulis</i>	Passion Kola	176.48 ± 0.25 ^A
<i>Dregea volubilis</i>	Aguna Kola	250.47 ± 0.27
<i>Solanum indicum</i>	Tibbatu	519.38 ± 5.87
<i>Solanum nigrum</i>	Kalukamberiya	268.47 ± 8.82
<i>Vernonia cinera</i>	Monarakudumbiya	>1000
<i>Acalypha indica</i>	Kuppameniya	267.86 ± 8.65
<i>Asparagus officinalis</i>	Hathawariya kola	253.27 ± 9.82
<i>Costus speciosus</i>	Tebu kola	>1000
<i>Sesbania grandiflora</i>	Kathurumurunga	519.38 ± 5.87
<i>Sapindus halicacabum</i>	Pennla	393.71 ± 4.63
<i>Amaranthus viridis</i>	Kura	591.38 ± 2.39
<i>Lasia spinosa</i>	Kohila dalu	738.08 ± 8.02
<i>Carum petroselinum</i>	Parsley leaves	>1000
<i>Coccinia grandis</i>	Kowakka kola	>1000
<i>Colocasia esculenta</i>	Kalualakola	358.99 ± 0.05
<i>Polyscias scutellaria</i>	Koppa kola	210.80 ± 0.17
<i>Lactuca sativa</i>	Salada	292.71 ± 3.02
<i>Sauropus androgynus</i>	Japanbatu kola	291.25 ± 2.33
<i>Coriandrum sativum</i>	Koththamalli kola	440.95 ± 5.32
<i>Cassia auriculata</i>	Ranawara	513.75 ± 5.06
<i>Mentha viridis</i>	Minchi	387.18 ± 2.96
<i>Aerva lanata</i>	Polpala	77.95 ± 0.62 **
<i>Baccopa monnieri</i>	Lunuwila	34.84 ± 0.13 **
Positive control	Quinoline	159.77 ± 1.45

Values are mean ± standard error (n=3), One-way ANOVA with Fisher's comparison. Other than ^A-values all the values are significantly different (p<0.001) than the positive control. ** - values are significantly lower than the positive control. ^A- values are not significantly different than the positive control.

RESULTS

It is observed that, most of the tested plant extracts such as *Centella asiatica*, *Ipomoea aquatica*, *Sesbania grandiflora*, *Murraya koenigii*, *Basella alba*, *Cardiospermum halicacabum*, *Vernonia cinerea*, *Costus speciosus*, *Lasiacpinosa*, *Coriandrumsativam*, *Menthaviridis*, *Carum petroselinum*, *Amaranthus viridus* and several others (see table 1) have significantly higher ($p < 0.001$) LC_{50} than the positive control. LC_{50} of the water extracts of *Aerva lanata* and *Baccopa monnieri* were found to be significantly lower ($p < 0.001$) than the positive control. Water extracts of *Alternanthera sessilis* and *Passiflora edulis* showed LC_{50} values, which are not significantly different from the positive control.

Interestingly all the extracts and the positive control tocopherol gave yellow spots, showing neutralization of the dark purple free radical, indicating the presence of free radical scavenging compounds in the plant extracts.

DISCUSSION

When LC_{50} values of all the leafy vegetables were analyzed using one-way ANOVA and Fisher's comparison with the positive control, other than *A. sessilis*, *P. edulis*, *A. lanata* and *B. monnieri* rest of the extracts were found to be significantly ($p < 0.001$) higher than the positive control (Quinoline), indicating their non cytotoxicity. It is observed that the LC_{50} values of *A. lanata* and *B. monnieri* are significantly lower than the positive control, indicating their higher cytotoxicity. Further, it is noted that according to the LC_{50} values, *A. sessilis* and *P. edulis* are equally toxic as the positive control to the Brine shrimps. Therefore, despite the fact that the majority of the extracts which have shown significantly higher LC_{50} values are non toxic, one can argue that consumption of *A. sessilis*, *P. edulis*, *A. lanata* and *B. monnieri* could lead to potential health risks.

According to the house-hold income and expenditure survey of Sri Lanka (2004) *A. sessilis*, locally known as Mukunuwenna / Ponankani, is consumed widely among Sri Lankans. That is mainly because of its taste and comparatively low costs; and large quantities are often being consumed in a single meal. Therefore, there is an important need for more investigations on toxicological aspects to evaluate their potential health risks to human beings. As the literature on toxicological studies of *A. sessilis* is inadequate, further toxicity studies should be carried out using cell lines and *in vivo* models to investigate their potential health risks. However, Broccoli

(*Brassica oleracea* var. *italica*), a very popular vegetable in the Western world is also claimed to be cytotoxic⁵. Yet no health warnings to the general public have been imposed by the respective authorities of these countries regarding eating Broccoli. Accordingly, consumption of fair amounts of *A. sessilis* may not possess health risks to human. Toxicity of *A. lanata*⁶, *B. monnieri*⁷ and *P. edulis*⁸ are previously reported, and our results are comparable with the literature data. However, in Sri Lanka *A. lanata* and *B. monnieri* are not consumed widely as leafy vegetables although used in traditional medicine.

Free radical scavenging properties demonstrate the presence of primary antioxidants in the plant extracts, which are claimed to be responsible for the chain breaking and free radical scavenging in body tissues⁴. The above observed antioxidant activity of the water extracts of common vegetable greens consumed in Sri Lanka, is a clear positive indication of the selection of appropriate green leafy vegetables by the local population.

Acknowledgement

The authors thank Dr G. Senaviratne for his valuable advice on statistical analysis and Ms. R. S. M. Perera and Mr. D. S. Jayaweera for technical assistance.

References

1. Lee T-H., Chen Y-M. & Chou H-N. (1999). Toxicity of cyanobacterial strains using *Artemia salina* in comparison with the mouse bioassay. *Acta Zoologica Taiwanica* **10**(1):1-8.
2. Michael A.S., Thompson C.G. & Abramovitz M. (1956). *Artemia salina* as a test organism for bioassay. *Science* **123**: 464.
3. Carball J.L., Hernandez-Indal Z.L., Perezl P. & Garcia-Gravalo M.D. (2002). A comparison between two brine shrimp assays to detect *in vitro* cytotoxicity of marine natural products. *BMC Biotechnology* **2**: 17-28.
4. Mattei R. (1998). Guarana (*Paullina cupana*): toxic behavioral effects in laboratory animals and antioxidants activity *in vitro*. *Journal of Ethnopharmacology* **60**(2): 111-116.
5. Martinez A., Ikken Y., Cambero M.I., Marin M.L., Haza A.I., Casas C. & Morales P. (1999). Mutagenicity and cytotoxicity of fruits and vegetables evaluated by the Ames test and 3-(4,5-dimethylthiazol-2-yl)2,5-diphenyltetrazolium bromide (MTT) assay. *Food Science and Technology International* **5** (5): 431-437.
6. Chowdhury D., Sayeed A., Islam A., Shah Alam Bhuiyan M. & Astaq Mohal Khan G.R.M. (2002). Antimicrobial activity and cytotoxicity of *Aerva lanata*. *Fitoterapia* **73**: 92-94.

7. D'Souza P., Deepak M., Rani P., Kadamboor S., Mathew A., Chandrashekar A.P. & Agarwal A. (2002). Brine shrimp lethality assay of *Bacopa monnieri*. *Phytotherapy Research* **16**(2):197-198.
8. Spencer K.C. & Seigler D.S. (1983). Cyanogenesis of *Passiflora edulis*. *Journal of Agricultural and Food Chemistry* **31**:794-796.